## ORCHID

# <u>Chromosomal Conditions Resolved With Whole Genome Sequencing (WGS) of Embryos</u>

Katz M<sup>1</sup>, Feinman<sup>1</sup>, Suer F<sup>1</sup>, Girod R<sup>1</sup>, Chandramohan D<sup>1</sup>, Li S<sup>1</sup>, Podgursky B<sup>1</sup>, Siddiqui N<sup>1</sup>, Xia Y<sup>1</sup> | Orchid Health, Durham, North Carolina

### BACKGROUND

As preimplantation genetic testing (PGT) technologies advance, so too does their resolution and ability to identify conditions that were previously undetectable<sup>1</sup>. High-depth whole genome sequencing (WGS now provides the capability to screen for chromosomal abnormalities missed by standard PGT for aneuploidy, including polyploidy, complete uniparental isodisomy (UPD, molar pregnancy), and microdeletions/ microduplications<sup>2</sup>. Embryos carrying such chromosomal abnormalities that are incorrectly classified as euploid may have serious health implications not only for the resulting fetus but also for the individual carrying the pregnancy<sup>3</sup>.

#### **PURPOSE & OBJECTIVES**

To present case studies that demonstrate how PGT-WGS can identify chromosomal conditions often missed by traditional PGT-A. We showcase three cases where chromosomal abnormalities were identified that could be overlooked with conventional methods, as well as a fourth case that highlights the use of PGT-M for structural variants, which would typically be rejected by other labs.

### **MATERIALS & METHODS**

Embryos from 4 separate couples were biopsied by trained embryologists at their respective In-vitro Fertilization (IVF) centers and sent to Orchid laboratory for testing. Each embryo biopsy underwent whole genome amplification using a lab-developed method followed by low-pass whole genome sequencing on the Illumina platform. Embryos marked as euploid were further analyzed by 30x whole genome sequencing.

#### RESULTS

In the first case, nine embryos were biopsied, and five were classified as euploid and, therefore, further analyzed by 30x WGS. One of the five euploid embryos was identified as having genomewide UPD, consistent with a molar embryo. This finding was made by reviewing allele tracks without requiring parental samples.

In the second case, five embryo biopsies were received, with two identified as euploid and sent for further analysis. One of these embryos was identified as aneuploid: 69, XXX (triploidy).

The third couple had 14 embryos, of which 11 were identified as euploid. Further analysis revealed that 6 of these embryos had a ~1.5 Mb microduplication at Xp22.31. This duplication has been observed in the general population at a rate of approximately 0.32-0.41% and has been associated with various neurodevelopmental phenotypes, most commonly seizures and learning disabilities<sup>4,5</sup>. Maternal inheritance was suspected since all the affected embryos were male (46, XY). Further counseling uncovered the patient's personal history of seizures, suggesting that the mother also exhibited symptoms. Confirmatory testing for the mother was recommended, along with a referral to a medical geneticist for additional health recommendations.

Finally, a fourth couple aimed to screen for a ~412 kb duplication on chromosome 10, associated with split-hand/foot syndrome, carried by the affected male partner. Since the duplication was de novo, traditional PGT-M probe development was not feasible, and other labs rejected their case. Of the three embryos tested, the two euploid embryos were negative for chromosome 10 duplication.



**Fig. 1**:

C.) Reference euploid copy number plot



Fig. 2:

1.7 Mb microduplication at Xp22.31. Copy number plot and variant allele frequency for affected/unaffected trophectoderm biopsies

#### CONCLUSIONS

PGT-WGS enables the detection of chromosomal and structural variants that may be missed by traditional PGT-A, such as molar pregnancies and clinically significant microdeletions/duplications. Additionally, PGT-WGS offers the possibility of PGT-M for challenging structural variants without requiring additional informative familial samples and/or probe development. This provides an option for families who other labs might otherwise reject due to technical limitations.

#### **FINANCIAL SUPPORT**

All authors are employees at Orchid Health.

### REFERENCES

- Treff NR, Zimmerman RS. Advances in Preimplantation Genetic Testing for Monogenic Disease and Aneuploidy. Annu Rev Genomics Hum Genet. 2017;18:189-200. doi:10.1146/annurev-genom-091416-035508
- Xia Y, Katz M, Chandramohan D, et al. The first clinical validation of whole-genome screening on standard trophectoderm biopsies of preimplantation embryos. F S Rep. 2024;5(1):63-71. Published 2024 Jan 11. doi:10.1016/j.xfre.2024.01.001
- Zhou B, Anglin HP, Quaas AM. Molar pregnancy after in vitro fertilization with euploid single embryo transfer. F S Rep. 2021;2(2):146-149. Published 2021 Jan 22. doi:10.1016/j.xfre.2021.01.003
- Mario Brinciotti, Francesca Fioriello, Antonio Mittica, Laura Bernardini, Marina Goldoni, Maria Matricardi, Epilepsy phenotype in patients with Xp22.31 microduplication, Epilepsy & Behavior Case Reports, Volume 11, 2019, Pages 31-34, ISSN 2213-3232, https://doi.org/10.1016/j.ebcr.2018.10.004.
- Esplin ED, Li B, Slavotinek A, Novelli A, Battaglia A, Clark R, Curry C, Hudgins L. Nine patients with Xp22.31 microduplication, cognitive deficits, seizures, and talipes anomalies. Am J Med Genet A. 2014 Aug;164A(8):2097-103. doi: 10.1002/ ajmg.a.36598. Epub 2014 May 6. PMID: 24800990.

A.) Genome-wide uniparental isodisomy (molar) copy number plot, B.) Triploidy (69, XXX) copy number plot,